Peptide Synthesis in Fluorinated Alcohols Mixed with Proton Accepting Partners

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Abstract: 1,1,1,3,3,3-Hexafluoro-2-propanol is an excellent dissolver of protected oligopeptides but a barren *medium for the condensation reaction of peptides. The solvent, however, could be changed to help the reaction proceed by mixing with a proton accepting solvent such as N,N-dimethylformamide.*

The solution-phase-peptide-synthesis often suffers from the solubility problem. The **protected** oligopeptide tends to lose its solubility to the common reaction media, such as dichloromethane and N , N dimethylformamide (DMP), though it depends on the amino acid sequences. On the other hand, it is widely known that fluorinated alcohols, such as 2,2,2-trifluoroethanol (TFE) and 1,1,1,3,3,3-hexafluoro-2-propanol $(HFIP)$, are good solvents for poorly soluble protected oligopeptides.¹ The dissolving capacity of these alcohols is considered to owe to their strong hydrogen-bonding ability with carbonyl groups in peptide bonds. Accordingly, the utilization of TFE and HFIP has been examined in the solid-phase-synthesis (SPS) of a difficult-sequence peptide.² The mixed solvent of CH_2Cl_2 and HFIP (9/1, v/v) was effective to condition the peptidyl resin support. However, to our knowledge has not been reported the successful application of TFE or HFIP to the condensation of poorly soluble peptide fragments. Thus, aiming the application of HFIP to the solution-phase-peptide-synthesis (fragment condensation), we attempted to find out the appropriate reaction condition, in which the reactivity of HFIP to the reagents is suppessed.

Using the condensation reaction of Boc-Ala-OH (Bee, t-butyloxycarbonyl) and H-Ala-OBzl (Bzl, benxyl) by N,N-dicyclohexylcarbodiimide (DCC)³ as a model system,⁴ we have found out at first that TFE gave Boc-Ala-Ala-OBzl in moderate yield (73%) but HPIP did not (15%) (Entry 7 and 2 in Table 1, respectively). The reactivity of the OH group of HFIP may be responsible for the barren medium. Therefore, an aprotic polar organic solvent, DMF was examined for the effect to decrease the reactivity of HFIP to DCC. For the pairs of TFE/DMF and HFIP/DMF, the condensation efficiency was plotted to their ratios (Fig. 1). Obviously, the increasing amount of DMF to HPIP improved the yield more drastically than to TPE. The higher reactivity of HFIP toward DCC was supposed to be tamed by DMF with the hydrogen bond as illustrated in Fig. 2.

and DMF with the hydrogen bond.

Fig. 1. Condensation yields in various DMF contents in HFIP (O) and TFE (\bullet) .

The other aprotic polar and apolar solvents including dimethylsulfoxide (DMSO), pyridine, and CH_2Cl_2 were further examined for the capacity to protect the various condensation reagents. The results were summarized in Table 1. All the proton accepting solvents appeared to be effective to help DCC to produce Boc-Ala-Ala-OBzl in good yields (Entry 3-5 in Tablell. Pyridine, which is not a good solvent for poorly soluble protected peptides, gave the highest yield. On the contrary, CH_2Cl_2 was less effective in the condensation (Entry 6) probably due to its very weak proton accepting capacity. The effect of added solvents, in the case of TFE, was repeated similarly as observed in HFIP (Entry 8-11).

Various condensation reagents were also examined about the influences of HFIP and the coexisting effects of the proton accepting partners (Table 1). The addition of 1-hydroxybenzotriazole $(HOBt)^5$ to DCC increased the condensation efficiency (Entry 13). It is noteworthy that the benzotriazole-1-yl-oxytris(dimethylamino)phosphonium hexafluorophosphate (BOP)/HOBt method⁶ afforded 73% yield in HFIP alone (Entry 14). 'Ihe proton accepting DMF could improved it to 89% (Entry 15). Diphenylphosphoryl azide (DPPA)' appeared not to be a good reagent in the condensation in HFIP containing media (Entry 16 and 17). Milton and Milton described their experiences with HFIP not to recommend in the formation of HOBt active ester of N-protected amino acid with DCC.² However, in this study the DCC/HOBt method could be utilized for condensation in HFIP mixed with DMF.

It should be emphasized that HFIP is a powerful dissolver of protected oligopeptides. Therefore, the use of HFIP in the fragment condensation of poorly soluble peptides will bring the great benefit in the chemical production of valuable hormonal peptides. In this aspect, the practical usefulness depends on the freedom from the racemization. The racemization was tested by the newly developed system, in which Boc-Ala-Ala-OH is

coupled with H-Pro-NHCH₃ to give the diastereomeric tripeptide mixture. The condensation was carried out with the selected reagents and mixed solvents (Table 2). The corresponding diastereomers of Boc-Ala-Ala-Pro-NHCH₃ (LLL and LDL) are successfully separated by HPLC.8 The DCC/HOBt method in $HFIP/DMF$ (1/1, v/v) caused racemization as low as 3.9%. The mixed solvent of HFIP/DMF/pyridine $(2/1/1, v/v/v)$ also appeared as a good medium with 1.6% racemization. However, the BOP/HOBt method was judged to be unsuitable for the fragment condensation, because it afforded 12% and 7.2% racemization in DMF alone and HFIP/DMF (1/1, v/v), respectively.

Table 2. Racemization during the Condensation in HFIP Containing Media.

The utility of HFIP/DMF/pyridine $(2/1)$, $v/v/v$ with DCC/HOBt was further demonstrated in the condensation of Ac-Glu(OcHex)-Leu-Leu-Lys(ClZ)-Ala-Pya-Ala-Glu(OcHex)-Leu-Leu-Lys(ClZ)-OH (a model peptide for the amphiphilic α -helix structure; cHex, cyclohexyl; Pya, L-1-pyrenylalanine) with H-Ala-OBzl as follows. Though the acid component, 11-peptide, was hardly soluble in DMF, complete dissolution with HFIP was attained. 'Ihe addition of DMF to a clear solution of the **1 1-peptide** in HFIP resulted in partial gelation. This fact suggested that the hydrogen bonding of HFIP to the carbonyl group of the peptide was replaced to that of DMF. Therefore, some mixed solvents based on HFIP were examined for better dissolution. Finally, an equal volume mixture of DMF and pyridine was added to the HFIP solution of the 11-peptide. In the slightly turbid solution, the condensation reaction with H-Ala-OBzl afforded the product in 81% yield and >90% purity on HPLC. It was treated with anhydrous HF and analyzed by FAB-MS $(m/z 1512 (M + H)^+)$. Thus, HFIP was shown as a useful co-solvent in the condensation of the peptide hardly soluble in DMF alone.

'Ihe solubility problem in the synthesis of peptide triggered the development of solid-phase-synthesis (SPS). However, SPS is difficult to be applied for the large scale synthesis of valuable peptides. The use of HFIP (or TFE if enough) by combination with a proton accepting pattner as discovered in the present study may revive the solution-phase-synthesis.

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- 4. 'lhe condensation was carried out as follows. To a chilled solution of Boc-Ala-OH (1.0 mmol) and H-Ala-OBzl p -toluenesulfonate (1.0 mmol) in the appropriate solvent (2.0 ml) was added the appropriate condensation reagent (1.5 mmol) and required triethylamine (1.0 mmol for DCC, 2.5 mmol for DPPA, and 3.25 mmol for BOP). The reaction was allowed to proceed for 5 h at room temperature. The product, Boc-Ala-Ala-OBzl, was extracted with 50 ml of ethyl acetate. After the N, N-dicyclohezyhuea was filtered off if required, the solution was washed with 10% citric acid, 4% NaHCO₃, and water, successively, and dried over MgSO₄. After the solvent was removed by evaporation, the residues were dissolved in methanoi (100 ml) and analyzed for Boc-Ala-Ala-OBzl by reversed-phase HPLC CYMC Cl8 column (4.6 x 150 mm) with a linear gradient of $10\n-100\%$ acetonitrile/0.1% trifluoroacetic acid over 30 min).
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- 8. HPLC was carried out on a YMC C18 column (4.6 x 150 mm) with a linear gradient of 10-100% acetonitrile/O.l% trifluoroacetic acid over 30 min (detection, 220 nm; flow rate, 1.0 ml/min). The retention times of LLL and LDL isomers were 12.1 and 13.2 min, respectively.

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